Breeding with molecular markers: a case study for tomato color and lycopene content

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Case study

Measuring traits, heritability and gain under selection

Developing markers for "candidate genes"

Estimating the proportion of phenotypic variance explained by a marker (related to concept of heritability)

Demonstrate where to apply markers in a breeding program: relative efficiency of selection

Select for candidate genes

Select for elite background "background selection".



Measuring traits: Color

- Color correlates with lycopene and beta carotene content. Beta carotene is a nutrient.
- Color uniformity determines product grade and grower premium.
- Color disorders (e.g. YSD) reduce color uniformity, price and nutritional quality.



Fresh-market

Processed

Plant Breeding is predominantly "phenotype" based
We select on the basis of traits
Growers buy varieties on the basis of trait performance
Measuring traits is therefore really important



Review: inbreeding drives individuals toward homozygosity.

A population of inbred lines can be replicated because seed from each plant in a line are identical.



Frequency of heterozygotes (Cc) and homozygotes (CC+cc) in each generation of selfing a hybrid (F1).

Measuring Traits

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Often a balance between accuracy and efficiency of data collection

Measuring traits: Objective measures of color



Phenotyping from digital images: fast and objective:

Tomato Analyzer (Brewer et al., *Plant Physiol*ogy, 2006, **141**, 15-25; Darrigues et al., JASHS, 2008, **133**, 579-586)









Colorimeter values

Analysis of color from Scanned images is consistent with colorimeter measurements and visual observations





Rank 216:

Rank 1:



Estimating heritability using variance estimates from ANOVA

Source	Expected MS
gen	Var(Error)+pVar(rep*gen)+qVar(year*gen)+(p-1)(q-1)Var(gen)
rep	Var(Error)+nVar(year*rep)+pVar(rep*gen) + (n)(p-1)Var(rep)
year	Var(Error)+nVar(year*rep)+qVar(year*gen) +(p-1)(q-1)Var(year)
year*gen	Var(Error) + q Var(year*gen)
rep*gen	Var(Error) + p Var(rep*gen)
year*rep	Var(Error) + n Var(year*rep)
gen*year*	rep Var(Error)

To estimate Var(Gen) use MS from ANOVA and solve. e.g. for 2 rep, 2 year trial:

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13.601 = Var(Error) + 2 Var(rep*gen) + 2 Var(year*gen) + 4 Var(gen)
2.6926 = Var(Error) + 2 Var(year*gen)
10.9084 = 2 Var(rep*gen) + 4 Var(gen)
3.4639 = 2 Var(rep*gen) + 2.77724
7.447 = 4 Var(gen) - 2.77724
10.22174 = 4 var(gen)
2.55 = var(gen) = \sigma^{2}(G)
h^{2} = \left[\sigma^{2}(G)\right] / \left[\sigma^{2}(G) + \sigma^{2}ge/e + \sigma^{2}error/r*e\right]
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Objective data from parents, offspring, and/or replicated trials allows us to estimate the genetic contribution to observed variation in a trait (heritability $h^2 = V_{additive genetic}/V_{total}$)

Expected and realized gain under selection is a function of heritability, the phenotypic variance in a population, and the selection differential (K) in standard deviation units.

Heritability can be estimated from parent-offspring regression or from variance partitioning (ANOVA)

Heritability for color increased with objective measurement.

Heritability for color increased when selection is based on a line or family across locations and years

Trait	individual	line or family
L	0.11	0.57
Hue	0.07	0.39



	Unselected	Selected
Average	7.41	3.71
STDEV	1.86	0.63

$$\Delta G = h^2(\sigma_P)K$$

Selection progress: reduction in within fruit variation (hue difference w/in fruit)



Take home message:

Score traits objectively on a quantitative scale whenever possible (improved measurement often means higher heritability).

Selection (K) is in standard deviation units relative to the population (σ_P).

Estimate heritability and realized gain at different stages of a breeding program.

Measure progress.





How can markers help make breeding more "efficient"?

- -Identifying markers
- -Using markers for selection

How do I identify markers linked to a trait?

Random approach (high density mapping).

"Candidate gene" approach.

DEFINITION of candidate gene:

A gene suspected of being involved in the expression of a trait.

Based on location

Based on presumed function (biochemical pathway, annotation, etc...)

Based on expression patterns



Carotenoid Biosynthesis: Candidate pathway for genes that affect color and color uniformity.



Disclaimer: this is not the only candidate pathway...

Neoxanthin

Databases that link pathways to genes

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Databases that link pathways to genes

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External Plant Metabolic databases

CapCyc (Pepper) (C. anuum) CoffeaCyc (Coffee) (C. canephora) SolCyc (Tomato) (S. lycopersicum) NicotianaCyc (Tobacco) (N. tabacum) PetuniaCyc (Petunia) (P. hybrida) PotatoCyc (Potato) (S. tuberosum) SolaCyc (Eggplant) (S. melongena)

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Using distributed resources (MetaCyc, NCBI, BLAST) we identified candidate genes, chromosomal locations, and polymorphisms.

Zeaxanthin epoxidase

Probable location on Chromosome 2

Alignment of Z83835 and EF581828 reveals 5 SNPs over ~2000 bp

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Comment on the databases:

Information is not always complete/up to date.

Display is not always optimal, and several steps may be needed to go from pathway > gene > potential marker.

s validated markers.



Germplasm data may also have error (e.g. PI 128216)

There is a wealth of information organized and available.

The previous example detailed how we might identify sequence based markers for trait selection.

Improving efficiency of selection in terms of 1) relative efficiency of selection, 2) time, 3) gain under selection and 4) cost will benefit from markers for both forward and background selection.

Remainder of Presentation will focus on

Where to apply markers in a program

- Forward and background selection
- Marker resources
- Alternative population structures and size

Establishing linkage between a "marker" and "trait"





Analysis is conceptually simple (t-test; ANOVA; regression)

In practice, more complex statistical models are adopted such that missing data is tolerated, significance level is accurately estimated, etc...

Example: QTL for color uniformity in elite crosses



How much phenotypic variation is explained by a QTL?

Analogous to the concept of "heritability" (variation due to genetics/total variation).



 r^2 for marker is dependent on two things: the strength of the linked gene and linkage distance. Therefore r^2 can be increased by identifying markers closer to the QTL or gene of interest.

Source	DF	Expected MS
Genotypes	N-1	σ2 + bσ2(G)
Marker	1	$\sigma^2 + b[\sigma^2(G_{QTL}) + 4r(1-r)g^2] + bc(1-2r)^2g^2$
Gen(marker)	N-2	$\sigma^2 + b[\sigma^2(G_{QTL}) + 4r(1-r)g^2]$
Error	N(b-1)	σ2

b is the number of replicatesr is the recombination separating the marker from the QTLc is a coefficient related to the population size

 $c = N - (n_1^2 + n_2^2)/N$

 $(n_1 + n_2 = 1;$ representing the number in each marker class) g is the genetic effect (in BC pop's additive and dominance effects are confounded).

 $\sigma 2(G_{QTL})$ is the part of the error variance that cannot be explained by the QTL.

V_{marker}/V_{total}

F-test for significance is Marker/Gen(marker)

 $= bc(1 - 2r)^2 g^2$

Significance depends on population size, recombination, the strength of the genetic effect relative to the error variance and the part of the error variance that cannot be explained by the QTL The proportion of variance explained by polymorphic marker on chromosome 2

Trait	$\underline{V}_{\underline{m}}/\underline{V}_{\underline{T}}$
L	0.25
Hue	0.15
L uniformity	0.28
Hue uniformity	0.32

Marker assisted selection is a form of indirect selection



MAS is a form of indirect selection: The relative efficiency of selection can be expressed as = $r(gen) \{H_{indirect}/H_{trait}\}$

H_{indirect} marker or indirect phenotype (proportion of accurate genotypes when scoring a marker)

H_{trait} Heritability of the trait

 $r_{(gen)}$ Genetic covariance for traits.

 $r_{(gen)}$ for a marker and a phenotypic trait = Var(marker)/Var(Total) or $r_{(marker-trait)}$

Comparison of direct selection with indirect selection (MAS).

Line-mean heritab	ility (H) ⁻	for colo						
H H					Indirect Selection			
Trait	plant	SE	line	SE	Prop Vp SE			
BRIX	0.14	0.09	0.40	0.26				
Color-L	0.11	0.04	0.57	0.22	L-MAS	0.25	0.15	
Color-Hue	0.07	0.04	0.39	0.23	Hue-MAS	0.15	0.06	
Color unifL	0.14	0.11	0.63	0.25	Ldiff-MAS	0.28	0.16	
Color unifHue	0.13	0.10	0.64	0.23	Hdiff-MAS	0.32	0.14	

Relative efficiency of selection: $r_{(gen)} \ge \{H_i/H_d\}$ Line performance over locations > MAS > Single plant

 $r_{(gen)} \propto \{H_i/H_d\}$

For Hue uniformity, when applied to early generations: = 0.32/0.13 = 2.46later generations: = 0.32/0.64 = 0.5

The best time to use genetic markers : early generation selection



Frequency distribution: Proportion of recurrent parent genome in BC1 population



Two-stage selectio	n		
Select for target	Select for RP genome at unlinked markers		
Three-stage select	ion	l	
Select for target allele	Select for RP recombinants at flanking markers	Select for RP genome at unlinked markers	
Four-stage selection)n		
Select for target	Select for RP recombinants at flanking	Select for RP genome on carrier	Select for RP genome at unlinked
allele	markers	chromosome	markers

References:

Frisch, M., M. Bohn, and A.E. Melchinger. 1999. Comparison of Selection Strategies for Marker-Assisted Backcrossing of a Gene. Crop Science 39: 1295-1301. For effective background selection we need:

Markers for our target locus (C > T SNP for Zep)

Markers on the target chromosome (Chrom. 2)

Markers unlinked to the target chromosome (~2 per chromosome arm)

Q10 of RP genome in percent									
	Population Size								
	20	40	60	80	100	125	150	200	
Two-Stage									
BC1	76.7	78.7	79.7	80.3	80.7	81.3	81.7	82.2	
BC2	90.3	91.9	92.8	93.3	93.6	93.9	94.0	94.6	
BC3	95.8	96.2	97.1	97.3	97.4	97.5	97.6	97.8	
Three-Stage									
BC1	71.2	72.7	73.4	73.6	73.3	73.2	72.8	72.2	
BC2	86.1	87.2	88.5	89.3	90.2	90.7	91.3	91.8	
BC3	94.4	95.7	96.5	96.9	97.2	97.3	97.5	97.6	

Q10 indicates a 90% probability of success From Frisch et al., 1999.

Important point: we can save ~1 generation in a BC program through background selection

The best time to use genetic markers : early generation selection

Selecting for recurrent parent genome can save >1 BC

Restructuring breeding programs to maximize use of MAS may include:

1) Increasing genotypic replication (population size) at the expense of plot replication (consider augmented designs).

2) Collecting and archiving objective data.

3) Restructuring selection pipelines to make optimal use of genotyping reagents.

References:



Brewer et al., 2006. Plant Physiology 141: 15-25

Darrigues et al., 2008. JASHS, 133: 579-586

Frisch, et al., 1999. Crop Science 39: 1295-1301.

Knapp and Bridges, 1990. Genetics 126: 769-777.