DIFFERENT TYPES OF PCR

polymerase chain reaction (PCR):

- It is a molecular technology aim to amplify a single or few copies of the DNA to thousands or millions of copies.
- Developed in 1983 by Kary Mullis, PCR is now a common and often indispensable technique used in medical and biological research labs for a variety of applications. These include diagnosis of infectious diseases, DNA sequencing and DNA-based phylogeny.
- In 1993, Mullis was awarded the Nobel prize in Chemistry along with Michael Smith for his work on PCR.

Types of PCR ??

- Conventional (Qualitative) PCR.
- Multiplex PCR.
- Nested PCR.
- RT-PCR and qRT-PCR.
- Quantitative PCR.
- Hot-start PCR.
- Touchdown PCR.
- Assembly PCR.
- Colony PCR.
- Methylation-specific PCR.
- LAMP assay.



Multiplex-PCR:

It is a special type of the PCR used for detection of multiple pathogens by using Multiple primers sets each one targets a particular pathogen.

Uses:

This permits the simultaneous analysis of multiple targets in a single sample.



Nested-PCR:

- Used to increase the specificity of DNA amplification.
- Two sets of primers are used in two successive reactions.
- In the first PCR, one pair of primers is used to generate DNA products, which will be the target for the second reaction.

Using one ('hemi-nesting') or two different primers whose binding sites are located (nested) within the first set, thus increasing specificity.

Uses:

Detection of pathogens that occur with very few amount.



RT-PCR (*Reverse Transcription PCR*, *Real Time - PCR*)

- Used to reverse-transcribe and amplify RNA to cDNA.
- PCR is preceded by a reaction using reverse transcriptase, an enzyme that converts RNA into cDNA.
- The two reactions may be combined in a tube.
 Uses:
- 1 Detection of RNA virus like (HCV).
- 2Detection of other M.O. through targeting of their Ribosomal RNA.

Reverse Transcription PCR, Real Time - PCR



Reverse Transcription PCR, Real Time - PCR



Quantitative Real-Time PCR (qRT-PCR)

 Method use fluorescent dyes, such as Sybr Green, or fluorescence-containing DNA probes, such as TaqMan, to measure the amount of amplified product as the amplification progresses. Progress of **DNA** amplification during *real time* (**RT-PCR**) by measuring the release of fluorescent "flashes" during amplification. A computer measures the rate of "flashing" in 96 simultaneous **experimental PCR** reactions relative to a **control** reaction



Quantitative – PCR:

- Used to measure the specific amount of target DNA (or RNA) in asample.
- By measuring amplification only within the phase of true exponential increase, the amount of measured product more accurately reflects the initial amount of target.
- Special thermal cyclers are used that monitor the amount of product during the amplification.

Hot-start PCR:

It is a technique performed manually by heating the reaction components to the DNA melting temperature (e.g. 95°C) before adding the polymerase.

Touchdown PCR:

- In this type the annealing temperature is gradually decreased in later cycles.
- ➤ The annealing temperature in the early cycles is usually 3-5°C above the standard T_m of the primers used, while in the later cycles it is a similar amount below the T_m.
- The initial higher annealing temperature leads to greater specificity for primer binding, while the lower temperatures permit more efficient amplification at the end of the reaction.

Assembly-PCR

(also known as Polymerase Cycling Assembly or PCA)

- In this type synthesis of long DNA structures by performing PCR on a pool of long oligonucleotides with short overlapping segments, to assemble two or more pieces of DNA into one piece.
- It involves an initial PCR with primers that have an overlap and a second PCR using the products as the template that generates the final full-length product.
- This technique may substitute for Ligation-based assembly

Assembly PCR:



Assembly PCR:





Bacterial colonies are screened directly by PCR, for example, the screen for correct DNAvector constructs.

Colonies are sampled with a sterile pipette tip and a small quantity of cells transferred into a PCR mix.

Methylation-specific PCR (MSP)

- Used to identify patterns of DNA methylation at cytosine guanine islands (C&G islands) in genomic DNA. CpG islands, are concerned in regulation of gene expression in mammalian cells.
- Target DNA is first treated with sodium bisulfite, which converts unmethylated cytosine bases to uracil, which is complementary to adenosine in PCR primers.
- > Two amplifications are then carried out on the bisulfite-treated DNA:

Methylation-specific PCR (MSP)

- One primer set anneals to DNA with cytosine (corresponding to methylated cytosine),
 The other set anneals to DNA with uracil
- (corresponding to unmethylated cytosine).
- MSP used in quantitative PCR provides quantitative information about the methylation state of a given CpG island.

Methylation-specific PCR (MSP)



LAMP assay: (Loop-mediated isothermal amplification)

- It is a Modified type of the PCR using 3:6 primers sets one of them is loop like primer.
- This test use Bst-polymerase enzyme
- (Bacillus stearothermophilus DNA Polymerase).
- Using only two temperatures (63°C for 45 min. then 85°C for 5 min.), may be carry out in water path.

Before you speak: THIR

T: is it True? H: is it Helpfull? I: is it Inspiring? N: is it Necessary K: is it Kind?